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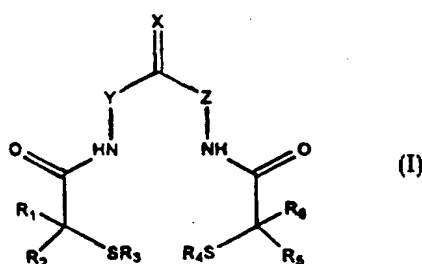


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(54) Title: HYDRAZINO-TYPE N<sub>2</sub>S<sub>2</sub> CHELATORS



(57) Abstract

Radionuclide chelating compounds are provided for conjugation to targeting molecules such as proteins, peptides or antibodies. The resulting radiolabelled compounds are useful in diagnosis and therapy. In formula (I) R<sub>1</sub>, R<sub>2</sub>, R<sub>5</sub> and R<sub>6</sub> are independently selected from H; carboxyl; C<sub>1-3</sub>alkyl; and C<sub>1-3</sub>alkyl substituted with a group selected from hydroxyl, sulfhydryl, halogen, carboxyl and aminocarbonyl; R<sub>3</sub> and R<sub>4</sub> are independently selected from H and a sulfur protecting group; X is selected from O, the group NH<sub>2</sub><sup>+</sup>, and the group CH<sub>2</sub>; Y and Z are independently selected from the group CR<sub>1</sub>R<sub>2</sub> and NR<sub>7</sub>; and R<sub>7</sub> is selected from H, carboxyl, C<sub>1-3</sub>alkyl and C<sub>1-3</sub>alkyl substituted with a group selected from hydroxyl, carboxyl and halogen.

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HYDRAZINO-TYPE  $N_2S_2$  CHELATORSField of the Invention

This invention is in the field of diagnostic imaging, and relates to chemical chelators useful in the radiolabelling of agents that target tissues of diagnostic interest.

5

Background to the Invention

The art of diagnostic imaging exploits contrasting agents that in binding or localizing site selectively within the body, help to resolve the image of diagnostic interest.  $^{67}$ Gallium-citrate, for example, has affinity for tumours and infected tissue and, with the aid of scanning tomography, can reveal afflicted body regions to the physician. Other contrasting agents include the metal radionuclides such as  $^{99m}$ technetium and  $^{186/188}$ rhenium, and these have been used to label targetting molecules, such as proteins, peptides and antibodies that localize at desired regions of the human body.

As targetting agents, proteins and other macromolecules can offer the tissue specificity required for diagnostic accuracy; yet labelling of these agents with metal radionuclides is made difficult by their physical structure. Particularly, protein and peptide targetting agents present numerous sites at which radionuclide binding can occur, resulting in a product that is labelled heterogeneously. Also, and despite their possibly large size, proteins rarely present the structural configuration most appropriate for high affinity radionuclide binding, i.e. a region incorporating four or more donor atoms that form five-membered rings. As a result, radionuclides are bound typically at the more abundant low-affinity sites, forming unstable complexes.

To deal with the problem of background binding, Paik et al (Nucl Med Biol 1985, 12:3) proposed a method whereby labelling of antibody is performed in the presence of excess DPTA (diaminetrimethylenepentaacetic acid), to mask the low affinity binding sites. While the problem of low affinity binding is alleviated by this method, actual binding of the radionuclide, in this case technetium, was consequently also very low.

The direct labelling of proteins having a high proportion of cysteine residues also has

been demonstrated (Dean et al; WO 92/13,572). This approach exploits thiol groups of cysteine residues as high-affinity sites for radionuclide binding, and is necessarily limited in application to those targeting agents having the required thiol structure.

5 A promising alternative to the direct labelling of targeting agents is an indirect approach, in which targeting agent and radionuclide are conjugated using a chelating agent. Candidates for use as chelators are those compounds that bind tightly to the chosen metal radionuclide and also have a reactive functional group for conjugation with the targeting molecule. For use in labelling peptide and protein-based targeting  
10 agents, the chelator is ideally also peptide-based, so that the chelator/targeting agent conjugate can be synthesized in toto using peptide synthesis techniques. For utility in diagnostic imaging, the chelator desirably has characteristics appropriate for its *in vivo* use, such as blood and renal clearance and extravascular diffusibility.

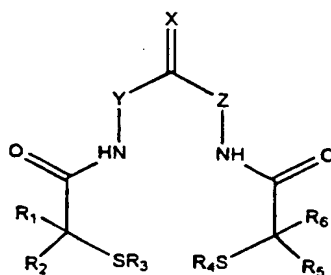
15 Summary of the Invention

The present invention provides chelators that bind diagnostically and therapeutically useful metal radionuclides, and can be conjugated to targeting agents capable of localizing at body sites of diagnostic and therapeutic interest. The chelators of the present invention are peptide analogues designed structurally to present an N<sub>2</sub>S<sub>2</sub>  
20 configuration capable of binding oxo, dioxo and nitrido ions of <sup>99m</sup>technetium and <sup>186/188</sup>renewium.

More particularly, and according to one aspect of the invention, there are provided metal radionuclide chelators of the formula:

25

(II)



wherein

$R_1$ ,  $R_2$ ,  $R_5$  and  $R_8$  are independently selected from H; carboxyl;  $C_{1-3}$  alkyl; and  $C_{1-3}$  alkyl substituted with a group selected from hydroxyl, sulfhydryl, halogen, carboxyl and aminocarbonyl;

- 5  $R_3$  and  $R_4$  are independently selected from H and a sulfur protecting group;  
X is selected from O, the group  $NH_2^+$ , and the group  $CH_2$ ;  
Y and Z are independently selected from the group  $CR_1R_2$  and  $NR_7$ ; and  
 $R_7$  is selected from H, carboxyl,  $C_{1-3}$  alkyl and  $C_{1-3}$  alkyl substituted with a group  
selected from hydroxyl, carboxyl and halogen.

10

According to another aspect of the invention, the chelators of the above formula are provided in a form having a metal radionuclide complexed therewith.

15

In another aspect of the invention, the chelator is provided in a form having a conjugating group attached thereto for coupling to a diagnostically useful targetting molecule, and optionally in combination with a complexed metal radionuclide, for imaging use.

#### Detailed Description of the Invention

20

The invention provides metal radionuclide chelators that when complexed with a radionuclide and conjugated to a targetting molecule are useful for delivering the radionuclide to a body site of therapeutic or diagnostic interest. As illustrated in the above formula, the chelators have an  $N_2S_2$  configuration in which the radionuclide is complexed.

25

Terms defining the variables  $R_1$  -  $R_7$  as used hereinabove have the following meanings:

"alkyl" refers to a straight or branched  $C_1$ - $C_8$  chain and embraces the term "lower alkyl" which refers to a straight or branched  $C_1$ - $C_3$  chain;

30

"halogen" refers to F, Cl, Br and I;

"sulfur protecting group" refers to a chemical group that inhibits oxidation, in particular those that are cleaved upon chelation of the metal. Sulfur protecting groups include known alkyl, aryl, acyl, alkanoyl, aryloyl, mercaptoacyl and organothio groups.

5

In preferred embodiments of the invention, the chelators conform to the above formula in which:

10  $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are independently selected from H and a lower alkyl group selected from ethyl and propyl; and most preferably are all H;

$R_3$  and  $R_4$  are a hydrogen atom or a sulfur protecting group selected from benzoyl, acetamidomethyl and substituted or unsubstituted tetrahydropyranyl groups;

X is selected from O, the group  $\text{CH}_2$ , and most preferably the group  $\text{NH}_2^+$ ;

15 Y and Z are independently selected from the groups  $\text{CR}_1\text{R}_2$  and  $\text{NR}_7$ , wherein at least one and most preferably both of Y and Z is  $\text{NR}_7$ ; and

$R_7$  is selected from carboxyl, lower alkyl and most preferably hydrogen.

20 Chelators having achiral carbon centres will advantageously alleviate the problems associated with differing biodistribution of various stereoisomers. In order to maintain achirality, a conjugating group must be attached to chelators of the invention at X. Attachment of a conjugating group at X is most stable when X is the group  $\text{NH}_2^+$ . Accordingly, it is a preferred embodiment of the present invention to provide chelators having achiral carbon centres and have a conjugating group attached to an  $\text{NH}_2^+$  group at X.

25

In specific embodiments of the invention, the chelators conform to the above general formula wherein  $R_1$  is H or a sulfur protecting group;  $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are each H; X is O,  $\text{CH}_2$  or  $\text{NH}_2^+$ ; and Y and Z are  $\text{CH}_2$  or NH. Specific examples include:

N,N'-bis-(S-benzoylmercaptoacetyl)-carbohydrazide;

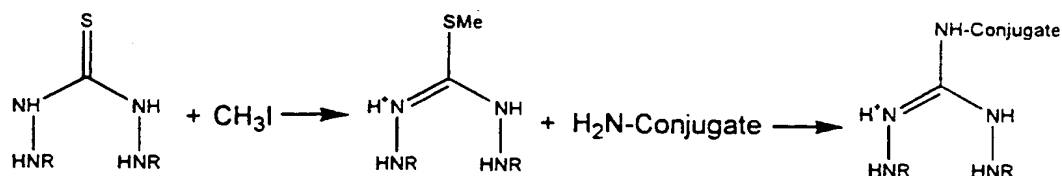
30 N,N'-bis-(S-Benzoylmercaptoacetyl)-1,3-diaminoguanidine hydrochloride; and



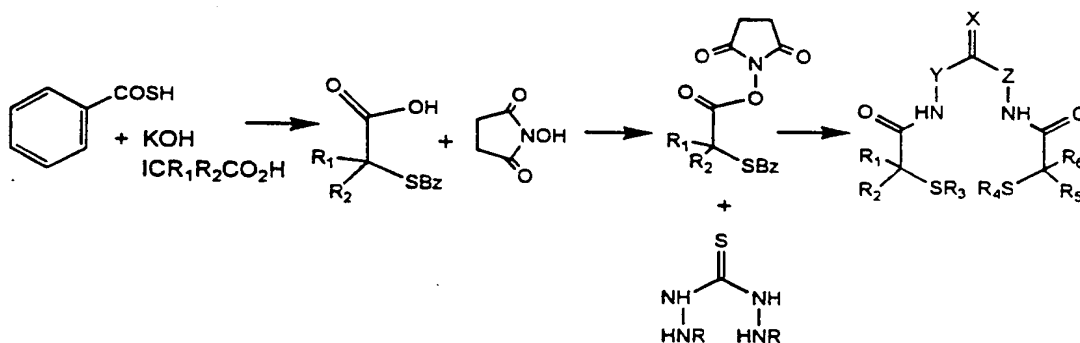
N,N'-bis-(S-benzoylmercaptoacetyl)-1,3-diaminoacetone.

For coupling to a targetting molecule, X or one of R<sub>1</sub>, R<sub>2</sub>, R<sub>5</sub> and R<sub>6</sub> desirably incorporates a "conjugating group", a term used hereinafter to refer to chemically reactive groups that allow for coupling of the chelator to a targetting molecule. In the preferred case where the targetting molecule is a peptide or protein, the conjugating group is reactive under conditions that do not denature or otherwise adversely affect the peptide or protein. In one embodiment of the invention, the conjugating group is reactive with a functional group of the peptide/protein such as an amino terminal group or an ε-amino group of a lysine residue so that the reaction can be conducted in a substantially aqueous solution. Useful conjugating groups include but are not limited to carboxyl groups, activated esters, a carboxy-methyl thio group, thiocyanates, amines, hydrazines, maleimides, thiols, and activated halides. In a preferred embodiment of the invention, conjugating groups are selected from methyl propanoate, carboxyl group and N-hydroxysuccinimide ester. Carboxyl conjugating groups may be activated with carbodiimide and an alcohol to form an active ester that is reactive with an amino group available on such targetting molecules as peptides and amino sugars, to form an amide linkage between the targetting molecule and the chelator conjugating group.

In certain embodiments, chelators of the present invention are provided wherein X is a CH<sub>2</sub> or oxo group. In a preferred embodiment, X forms an NH<sub>2</sub><sup>+</sup> group that may have a conjugating group attached thereto forming the group NH<sup>+</sup>-conjugate. A conjugating group may be introduced to this type of chelator during standard synthesis of a diaminoguanidine intermediate. Briefly, thiocarbohydrazine is converted to methyl-thiocarbohydrazine by substitution with methyl iodide. Addition of a selected amino-substituted conjugating group to the methyl-thiocarbohydrazine results in an intermediate which is subsequently employed in the preparation of chelators wherein X is NH<sub>2</sub><sup>+</sup> having a conjugate bound thereto. A method of preparing a conjugate bound intermediate is represented below.

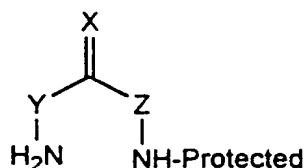


Chelators of the present invention that are symmetrical ie. in which  $R_1$  and  $R_2$  correspond to  $R_5$  and  $R_6$  respectively, can be prepared by the following general procedure. Commercially available iodoacetic acid, or a variant thereof substituted as desired, is reacted with potassium thiobenzoate yielding benzoylmercaptoacetic acid. This intermediate is then transformed into the corresponding N-hydroxysuccinimide ester using dicyclocarbodiimide in dioxane. The active ester is reacted with a selected carbonylhydrazide or diaminoguanidine or diaminoacetone yielding the symmetrical chelator. This method of preparing symmetrical chelators is illustrated below:



Preparation of chelators of the invention that are not symmetrical ie. in which  $R_1$  and  $R_2$  do not correspond to  $R_5$  and  $R_6$ , requires steps additional to those presented in the scheme above. In a particular method, benzoylmercaptoacetyl-N-hydroxysuccinimide, prepared as described above is reacted with a selected carbonylhydrazide,

diaminoguanidine or diaminoacetone of the general formula



The resulting benzoylmercaptoacetyl-carbohydrazide/-diaminoguanidine/  
-diaminoacetone is deprotected and subsequently reacted with a  
10 benzoylmercaptoacetyl-N-hydroxysuccinimide having the selected R<sub>5</sub> and R<sub>6</sub> groups  
to yield the asymmetrical N<sub>2</sub>S<sub>2</sub> chelator.

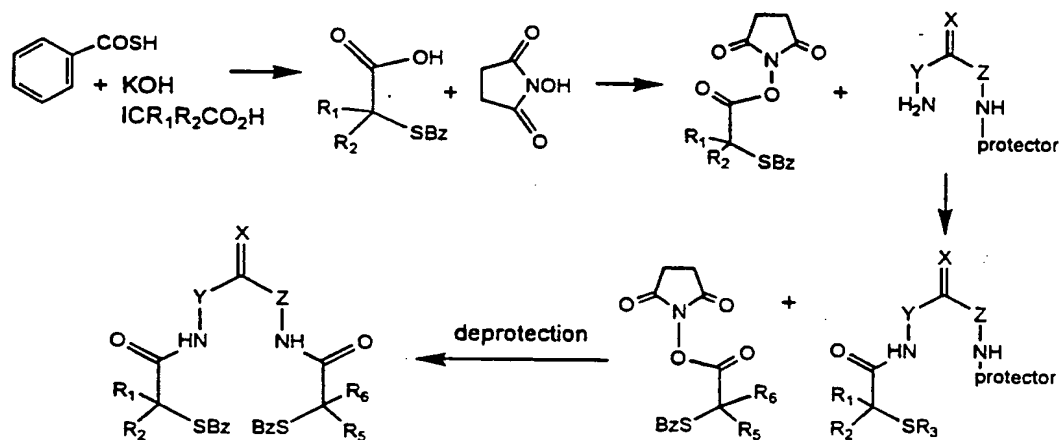
In a particular embodiment of the present invention, an N-protected diaminoguanidine  
is prepared from a hydrazine ester in which the OR group of the ester is substituted  
15 with an N-protected hydrazine to give the mono-N-protected diaminoguanidine.  
N-protecting groups common to the art may be used, for example *t*-butyloxycarbonyl  
(*t*-Boc) or 9-fluorenylmethyloxycarbonyl (Fmoc). Removal of *t*-Boc protecting groups  
can be achieved by addition of an anhydrous acid such as HCl in acetic acid while  
Fmoc may be cleaved with piperidine and diiodomethane or piperidine  
20 dimethylformamide.

Chelators of the present invention may also be asymmetrical with respect to Y and Z.  
That is, one of Y or Z may be the group CR<sub>1</sub>R<sub>2</sub> while the other is NR<sub>7</sub>. This type of  
asymmetry may be introduced by using the intermediate 1,3-diaminoamidine or a  
25 derivative thereof in place of the intermediate 1,3-diaminoguanidine in the synthetic  
processes previously described.

It is to be understood that variation at R<sub>1</sub>, R<sub>2</sub>, R<sub>5</sub> and R<sub>6</sub> can be introduced by using  
variants of iodo acetic acid. For example the alpha carbon may have one or two  
30 substituents as well as iodine such as lower alkyl, substituted lower alkyl or a

conjugating group. Variation at Y and Z may be introduced by using derivatives of carbohydrazide, diamino guanidine and diamino acetone. For example carbohydrazide and diamino guanidine intermediates may have carboxyl, lower alkyl, substituted lower alkyl or a conjugating group at either or both alpha nitrogens while diamino acetone may also have carboxyl; lower alkyl; substituted lower alkyl or a conjugating group at either or both alpha carbons.

A method of preparing asymmetric chelators of the invention is represented below:



For diagnostic imaging purposes, the chelators per se may be used in combination with a metal radionuclide. Suitable radionuclides include technetium and rhenium in their various forms such as <sup>99m</sup>TcO<sup>3+</sup>, <sup>99m</sup>TcO<sub>2</sub><sup>+</sup>, ReO<sup>3+</sup> and ReO<sub>2</sub><sup>+</sup>. Most desirably, and according to another aspect of the invention, the chelator is coupled through its conjugating group to a targeting molecule that serves to localize the chelated radionuclides at a desired site in a mammal. Examples of targeting molecules include, but are not limited to, steroids, proteins, peptides, antibodies, nucleotides and saccharides. Preferred targeting molecules include proteins and peptides, particularly those capable of binding with specificity to cell surface receptors characteristic of a

particular pathology. For instance, disease states associated with over-expression of particular protein receptors can be imaged by labelling that protein or a receptor binding fragment thereof in accordance with the present invention. Targetting peptides useful to image certain medical conditions and tissues are noted below:

5 for atherosclerotic plaque:

	YRALVDTLK	RALVDTLK
	RALVDTLK FVTQAEGAK	YAKFRETLEDTRDRMY
	AKFRETLEDTRDRMY	AALDLNAVANKIADFEL
	Y AALDLNAVANKIADFEL	YRALVDTLK FVTEQAKGA
10	RALVDTLK FVTEQAKGA	YRALVDTEFKVKQEAGAK
	RALVDTEFKVKQEAGAK	YRALVDTLK FVTQAEGAK

for infections and atherosclerotic plaque:

	VG VAPGVGVAPGVGVAPG	formyl.Nieu.LF.Nieu.YK
15	VPGVGVPGVGVPGVGVPGVG	formylIMIFL
	formylIMLFK	formylIMLFI
	formylIMFIL	formylIMFLI
	formylIMLIF	formylIMILF
	formylITKPR	VG VAPG
20	formylIMLF	YIGSR
	CH <sub>2</sub> CO.YIGSRC	

for thrombus:

	NDGDFEEIPEEYLQ	NDGDFEEIPEEY(SO <sub>3</sub> Na)LQ
25	GPRG	

for platelets:

	D-Phe.PRPGGGGNGDFEEIPEEYL	RRRRRRRRRGDV
	PLYKKIIKKLLES	RGD
30	RGDS	

for amyloid plaque (Alzheimer's disease):

EKPLQNFTLSFR

5 For connection to the chelator, a targetting molecule may comprise a "spacer" that serves to create a physical separation between the chelator and the targetting molecule. Suitable spacers are those that allow the targetting molecule to retain its *in vivo* localizing properties and are typically chemically inert groups such as an alkyl chain that is derivatized for coupling to the chelator. In the case where the targetting molecule is a peptide, the spacer may suitably be one or more amino acid residues. 10 Preferably, peptidic targetting molecules incorporate spacers of from 1 to 5 amino acids such as those having chemically inert  $\alpha$ -carbon side chains, such as glycine or  $\beta$ -alanine residues.

15 Peptide-based targetting molecules can be made using various established techniques. Because they are amenable to solid phase synthesis, alternating Fmoc protection and deprotection is the preferred method of making short peptides. Recombinant DNA technology is preferred for producing proteins and long fragments thereof. The chelators of the present invention can be coupled to a targetting molecule prior to labelling with a radionuclide, a process referred to as the "bifunctional chelate" 20 method. An alternative approach is the "prelabelled ligand" method in which the chelator is first labelled with a radionuclide and is then coupled to the targetting molecule.

25 Chelation of the selected radionuclide can be achieved by various methods. In a particular method, a chelator solution is first formed by dissolving the chelator optionally with a targetting molecule attached in aqueous alcohol eg. ethanol-water 1:1. The solution is degassed to remove oxygen then the thiol protecting groups are removed with a suitable reagent such as sodium hydroxide or by heating, and the resulting mixture is then neutralized with an organic acid such as acetic acid (pH 6.0- 30 6.5). In the chelation step, sodium pertechnetate is added to the chelator solution

with a reducing such as stannous chloride in an amount sufficient to reduce the technetium. The solution is mixed and left to react at room temperature and then heated on a water bath. In an alternative method, labelling can be accomplished with the chelator solution adjusted to pH 8. In this case, pertechnetate may be replaced with a solution of technetium complexed with ligands that are labile and exchangeable with the desired chelator. Suitable ligands include tartarate, citrate or heptagluconate. As a further alternative, sodium dithionite may be used as the reducing agent, and in this case the chelating solution is preferably adjusted to pH 12-13 for the labelling step. In all cases, the labelled chelator may be separated from contaminants  $^{99m}\text{TcO}_4^-$  and colloidal  $^{99m}\text{TcO}_2$  chromatographically, for example with a C-18 Sep Pak column activated with ethanol followed by dilute HCl. Eluting with dilute HCl separates the  $^{99m}\text{TcO}_4^-$ , and eluting with EtOH-saline 1:1 brings off the chelator while colloidal  $^{99m}\text{TcO}_2$  remains on the column.

Once prepared, the labelled chelator or chelator-targetting agent conjugate is administered to a patient by techniques well established in the art of radiodiagnostic imaging and radiotherapy. Typically a labelled chelator solution is administered by injection intravenously, intra-arterially, peritoneally or intratumorally in a pharmaceutically acceptable solution such as saline or blood plasma medium. The amount of labelled conjugate administered is dependent upon the toxicity profile of the chosen targetting molecule as well as the profile of the metal and is typically in the range of about 5-50 mCi/70 kg and more typically in the range of 25-35 mCi/70 kg. Localization of the metal *in vivo* is tracked by standard scintigraphic techniques at an appropriate time subsequent to its administration. The time at which an image may be obtained will depend in large part on the tissue distribution and clearance profile of the targetting molecule, for example most peptides will localize rapidly allowing for an image to be obtained within 3 hours and often within 1 hour.

The following examples are presented to illustrate certain embodiments of the present invention.

**Example 1 - PREPARATION OF N,N'-BIS-(S-BENZOYLMERCAPTOACETYL)-  
CARBOHYDRAZIDE RP-021**

To a purged stirring solution at 0°C of (5.74g, 41.6mmoles) thiobenzoic acid in (100mL) ethanol was added (27.8mL, 3N, 83.2mmoles) potassium hydroxide followed  
5 by (7.70g, 41.6mmoles) iodoacetic acid in (30mL) ethanol. The solution stirred for 10 hours at room temperature under argon. The ethanol was rotavapped and the orange solid product was dissolved in (40mL) water. The solution was acidified to pH 2.0 where an orange precipitate formed. The precipitate was filtered, washed with water, and dried in vacuo to give (7.98g, 99% yield) benzoylmercaptoacetic acid.

10

To a stirring solution of (9.20g, 46.9mmoles) benzoylmercaptoacetic acid and (5.41g, 46.9mmoles) N-hydroxysuccinimide in (100mL) dioxane was added a solution of (9.70g, 47mmoles) dicyclohexylcarbodiimide in (40mL) dioxane. The reaction was stirred 12 hours, followed by cooling to 4°C, filtering, and rotavapping off the dioxane  
15 to a white solid. The solid was triturated with cold isopropanol, filtered, and dried in vacuo yielding 10.8g, 78% benzoylmercaptoacetyl-N-hydroxysuccinimide. 200mg was recrystallized from 500mg in hot ethyl acetate.

20

To a stirring solution of (1.17g, 4.0mmoles) benzoylmercaptoacetyl-N-hydroxysuccinimide in (36mL) dioxane was added a solution of (180mg, 2.0mmoles) carbohydrazide in (10mL) dioxane. After 1 hour, TLC showed a partial reaction and (404mg, 4.0mmoles) triethylamine was added. The reaction stirred 4 hours followed by rotavapping off the dioxane and adding (5mL) water to a white solid. This was  
25 filtered, washed with (5mL) water, and dried in vacuo yielding 772mg, 86.5% N,N'-bis-(S-benzoylmercaptoacetyl)-carbohydrazide (m.p. 184-190 °C).

**Example 2 - PREPARATION OF N,N'-BIS-(S-BENZOYLMERCAPTOACETYL)-1,3-  
DIAMINOGUANIDINE HYDROCHLORIDE RP-032**

To a purged stirring solution at 0°C of (5.74g, 41.6mmoles) thiobenzoic acid in  
30 (100mL) ethanol was added (27.8mL, 3N, 83.2mmoles) potassium hydroxide followed



by (7.70g, 41.6mmoles) iodoacetic acid in (30mL) ethanol. The solution stirred for 10 hours at room temperature under argon. The ethanol was rotavapped and the orange solid product was dissolved in (40mL) water. The solution was acidified to pH 2.0 where an orange precipitate formed. The precipitate was filtered, washed with water, and dried in vacuo to give (7.98g, 99% yield) benzoylmercaptoacetic acid.

To a stirring solution of (9.20g, 46.9mmoles) benzoylmercaptoacetic acid and (5.41g, 46.9mmoles) N-hydroxysuccinimide in (100mL) dioxane was added a solution of (9.70g, 47mmoles) dicyclohexylcarbodiimide in (40mL) dioxane. The reaction was stirred 12 hours followed by cooling to 4°C, filtering, and rotavapping off the dioxane to a white solid. The solid was triturated with cold isopropanol, filtered, and dried in vacuo yielding 10.8g, 78% benzoylmercaptoacetyl-N-hydroxysuccinimide. 200mg was recrystallized from 500mg in hot ethyl acetate.

To a stirring solution of (200mg, 1.59mmoles) 1,3-diaminoguanidine•HCl in (3mL) methanol was added a solution of (936mg, 3.19mmoles) benzoylmercaptoacetyl-N-hydroxysuccinimide and (322mg, 3.19mmoles) triethylamine in (30mL) dioxane. The reaction stirred 14 hours overnight forming a fine white precipitate. The solvents were rotavapped off to a sticky white solid. The solid was triturated with cold isopropanol, filtered, washed with isopropanol, and dried in vacuo yielding 155mg, 32% N,N'-bis-(S-benzoylmercaptoacetyl)-1,3-diaminoguanidine hydrochloride (m.p. 126-128 °C).

**Example 3 - PREPARATION OF N,N'-BIS-(S-BENZOYLMERCAPTOACETYL)-1,3-DIAMINOACETONE RP-042**

To a purged stirring solution at 0°C of (5.74g, 41.6mmoles) thiobenzoic acid in (100mL) ethanol was added (27.8mL, 3N, 83.2mmoles) potassium hydroxide followed by (7.70g, 41.6mmoles) iodoacetic acid in (30mL) ethanol. The solution stirred for 10 hours at room temperature under argon. The ethanol was rotavapped and the orange solid product was dissolved in (40mL) water. The solution was acidified to pH 2.0 where an orange precipitate formed. The precipitate was filtered, washed with water,

and dried in vacuo to give (7.98g, 99% yield) benzoylmercaptoacetic acid.

To a stirring solution of (9.20g, 46.9mmoles) benzoylmercaptoacetic acid and (5.41g, 46.9mmoles) N-hydroxysuccinimide in (100mL) dioxane was added a solution of  
5 (9.70g, 47mmoles) dicyclohexylcarbodiimide in (40mL) dioxane. The reaction was stirred 12 hours followed by cooling to 4°C, filtering, and rotavapping off the dioxane to a white solid. The solid was triturated with cold isopropanol, filtered, and dried in vacuo yielding 10.8g, 78% benzoylmercaptoacetyl-N-hydroxysuccinimide. 200mg was recrystallized from 500mg in hot ethyl acetate.

10 To a stirring solution of (492g, 1.68mmoles) benzoylmercaptoacetyl-N-hydroxysuccinimide and (150mg, 0.84mmoles) 1,3-diaminoacetone hydrochloride monohydrate in (15mL) dioxane was added (850mg, 8.40mmoles) triethylamine. The reaction stirred 3.5 days followed by rotavapping off the dioxane to a yellow oil and  
15 adding (3mL) water to a yellow solid. The solid was crushed, filtered, washed with water, and dried in vacuo yielding 242mg, 65% N,N'-bis-(S-benzoylmercaptoacetyl)-1,3-diaminoacetone (m.p. 134-140 °C).

#### Example 4 - Preparation of <sup>99m</sup>Tc labelled chelators

20 Approximately 1mg of each chelator was dissolved in 200μL saline or 200μL ethanol:water (1:1) in a tube. To the tube was added 100-300μL sodium <sup>99m</sup>Tc-pertechnetate (5-15mCi), 100μL phosphate buffer (0.25M, pH 7.4), and 200μL of a solution containing 50μg stannous chloride dihydrate and 40mg sodium tartrate dihydrate. The tube was capped tightly and placed in a boiling water bath for 10  
25 minutes.

After cooling, the reaction mixture was loaded onto a C-18 solid-phase extraction cartridge (Sep-Pak) which had been activated with 5mL methanol and 5mL, 1mM HCl. The cartridge was washed with 5mM HCl and the eluate was collected in a test tube.

30 The cartridge was then dried by forcing air through it. The product was eluted with

2mL ethanol:water (1:1) and collected in a separate tube. The cartridge was placed in another tube and the three tubes were assayed in a radionuclide dose calibrator (ionization chamber). The yield was calculated as the activity in the ethanol:water eluate divided by the total activity in the three tubes under the assumption that the desired product was relatively lipophilic. A less lipophilic chelator would elute partially in the acid wash and the apparent yield would be low.

	Labelling yields		
	trial 1	trial 2	trial 3
Example 1	57%	59%	39%
Example 2	31%	25%	58%
Example 3	76%	75%	

Example 5 - In vivo distribution

Distribution within rats of the exemplified chelators and a reference chelator was determined using established protocols. Briefly, male Wistar rats (Charles River, 200g) were anaesthetized with somnitol (40 to 50mg/kg) and 200 $\mu$ L of the labelled chelator (ie. 200 $\mu$ Ci) was injected intravenously via the tail vein. Serial whole-body scintigrams were acquired after the first 10 minutes. Further images were obtained at 60 and 120 minutes, and then the rats were killed with anaesthesia and samples of organs (blood, heart, lung, liver, spleen, kidney, muscle, GI tract) were weighed and counted in either a well-type gamma counter or in a gamma dose calibrator depending on the organ. Dose calculations were made based on the assumption that rats weighed 200g and that the blood volume constituted 8% body weight. All results were corrected for the residual dose in the tail.

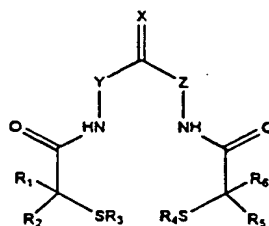
Each of the present chelators examined cleared relatively rapidly from the blood as

desired. For the chelator of example 1, it was found that the GI tract excretion was remarkably low in comparison with the  $N_2S_2$  reference chelator. The GI tract accounted for about 9% of the dose, and only about 14% remained in the blood. The chelator of example 2 localized primarily (30%) in the GI tract and (20%) in urine, with  
5 only about 10% of the dose remaining in the blood. The chelator of example 3 also localized primarily (50%) in the GI tract and (25%) in urine with only 3% in the blood.

Of these chelators, the chelator of example 3 showed the fastest clearance from the blood and other tissues and is mainly eliminated through the liver and GI tract. The  
10 chelators of example 1 and 2 had approximately equal accumulation in the kidney (7-14%).

WE CLAIM:

1. A compound of the general formula:

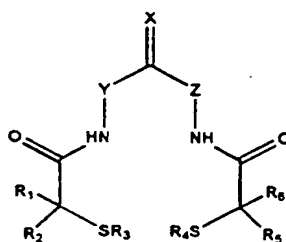


wherein

- $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are independently selected from H; carboxyl;  $C_{1-3}$  alkyl; and  $C_{1-3}$  alkyl substituted with a group selected from hydroxyl, sulfhydryl, halogen, carboxyl and aminocarbonyl;
- $R_3$  and  $R_4$  are independently selected from H and a sulfur protecting group;
- X is selected from O, the group  $NH_2^+$ , and the group  $CH_2$ ;
- Y and Z are independently selected from the group  $CR_1R_2$  and  $NR_7$ , wherein at least one of Y and Z is  $NR_7$ ; and
- $R_7$  is selected from H, carboxyl,  $C_{1-3}$  alkyl and  $C_{1-3}$  alkyl substituted with a group selected from hydroxyl, carboxyl and halogen.
2. A compound according to claim 1, wherein  $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are hydrogen.
3. A compound according to claim 1, wherein Y and Z are each the group  $NR_7$ .
4. A compound according to claim 3, wherein  $R_1$ ,  $R_2$ ,  $R_5$ ,  $R_6$  and  $R_7$  are hydrogen.
5. A compound according to claim 1, which is selected from:  
 $N,N'$ -bis-(S benzoylmercaptoacetyl)-carbohydrazide  
 $N,N'$ -bis-(S-Benzoylmercaptoacetyl)-1,3-diaminoguanidinehydrochloride
6. A compound according to claim 1, wherein  $R_3$  and  $R_4$  are selected from the

group consisting of a hydrogen atom, benzoyl group, an acetamidomethyl group and a substituted or unsubstituted tetrahydropyranyl group.

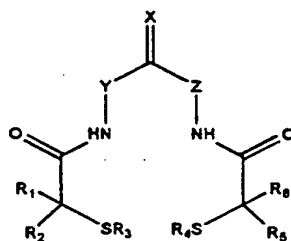
7. A compound according to claim 1, in a form complexed with a metal radionuclide or an oxide or nitride thereof.
8. A compound according to claim 1, in a form complexed with  $^{99m}\text{Tc}$  or oxide thereof.
9. A compound of the general formula:



wherein

- $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are independently selected from H; carboxyl;  $C_{1-3}$  alkyl;  $C_{1-3}$  alkyl substituted with a group selected from hydroxyl, sulfhydryl, halogen, carboxyl and aminocarbonyl; and a conjugating group for coupling a targeting molecule;
- $R_3$  and  $R_4$  are independently selected from H and a sulfur protecting group;
- X is selected from O, the group  $\text{NH}_2^+$  and the group  $\text{CH}_2$ , optionally having attached thereto a conjugating group for coupling a targeting molecule;
- Y and Z are independently selected from the group  $\text{CR}_1\text{R}_2$  and  $\text{NR}_7$ , wherein at least one of Y and Z is  $\text{NR}_7$ ; and
- $R_7$  is selected from H, carboxyl,  $C_{1-3}$  alkyl and  $C_{1-3}$  alkyl substituted with hydroxyl, carboxyl or halogen;
- wherein said compound incorporates a conjugating group at one of  $R_1$ ,  $R_2$ ,  $R_5$ ,  $R_6$  and X.

10. A compound according to claim 9, wherein the conjugating group is selected from the group consisting of carboxyl, N-hydroxysuccinimide ester and methyl propanoate.
11. A compound according to claim 9, wherein  $R_1$ ,  $R_2$ ,  $R_5$ , and  $R_6$  are hydrogen and X has a conjugating group attached thereto.
12. A compound according to claim 11, wherein X is  $\text{NH}_2^+$ .
13. A compound according to claim 9, in a form complexed with a metal radionuclide or an oxide or nitride thereof.
14. A compound according to claim 9, in a form complexed with  $^{99m}\text{Tc}$  or oxide thereof.
15. A compound of the general formula:



wherein

- $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are independently selected from H; carboxyl;  $\text{C}_{1-3}$  alkyl;  $\text{C}_{1-3}$  alkyl substituted with a group selected from hydroxyl, sulfhydryl, halogen, carboxyl and aminocarbonyl; and a conjugating group having a targetting molecule attached thereto;
- $R_3$  and  $R_4$  are independently selected from H and a sulfur protecting group;
- X is selected from O, the group  $\text{NH}_2^+$ , or the group  $\text{CH}_2$  each of which may have a targetting molecule attached thereto via a conjugating group;

Y and Z are independently selected from the group  $CR_1R_2$  and  $NR_7$ ; and  $R_7$  is selected from H, carboxyl,  $C_{1-3}$  alkyl and  $C_{1-3}$  alkyl substituted with hydroxyl, carboxyl or halogen;

wherein a conjugating group having a targetting molecule attached thereto is incorporated at one of  $R_1$ ,  $R_2$ ,  $R_5$ ,  $R_6$  or X.

wherein at least one of  $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  is a conjugating group having a targetting molecule attached thereto or X has a targetting molecule attached thereto via a conjugating group.

16. A compound according to claim 15, wherein  $R_1$ ,  $R_2$ ,  $R_5$  and  $R_6$  are H; and X is  $NH_2^+$  having a targetting molecule attached thereto via a conjugating group.
17. A compound according to claim 15, wherein the targetting molecule is a protein or peptide
18. A compound according to claim 15, in a form complexed with a metal radionuclide or an oxide or nitride thereof.
19. A compound according to claim 15, in a form complexed with  $^{99m}Tc$  or oxide thereof.



## INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 94/00479

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C07C327/32 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07C

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	JOURNAL OF MEDICINAL CHEMISTRY, vol.29, no.10, October 1986 Washington, DC, US, pages 1933 - 1940 S. KASINA, ET AL.: 'Tissue distribution properties of technetium-99m-diamide- dimercaptide complexes and potential use as renal radiopharmaceuticals' see table, ligand 21 --- -/--	1,9,15



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

## \* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
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- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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\*&\* document member of the same patent family

Date of the actual completion of the international search

25 October 1994

Date of mailing of the international search report

- 9. 11. 94

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English, R

# INTERNATIONAL SEARCH REPORT

International Application No  
PC1/CA 94/00479

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>JOURNAL OF NUCLEAR MEDICINE,, vol.24, no.2, February 1984 New York, US, pages 223 - 229 R.F. SCHNEIDER, ET AL.: 'N,N'-bis(S-Benz- oylmercaptoacetamido)ethylenediamine and propylenediamine ligands as renal function imaging agents. I. Alternative synthetic methods' see compound 27</p> <p style="text-align: center;">-----</p>	1,9,15